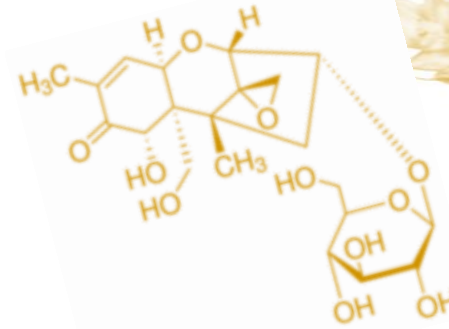
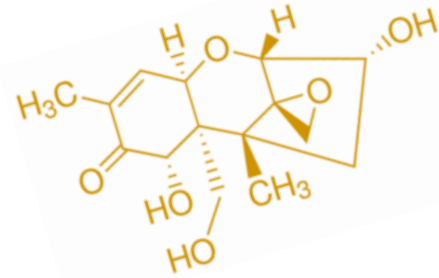


toxins

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Analysis of Deoxynivalenol and Deoxynivalenol-3-glucoside in Hard Red Spring Wheat Inoculated with *Fusarium graminearum*.

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Introduction



Fusarium Head Blight



Fungal pathogens cause the Fusarium Head Blight (FHB) disease in wheat and other cereal crops

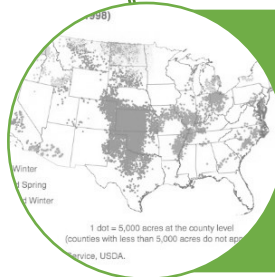
FHB as a worldwide problem



Fusarium



Global problem with outbreaks in the USA, Canada, Europe and Central America

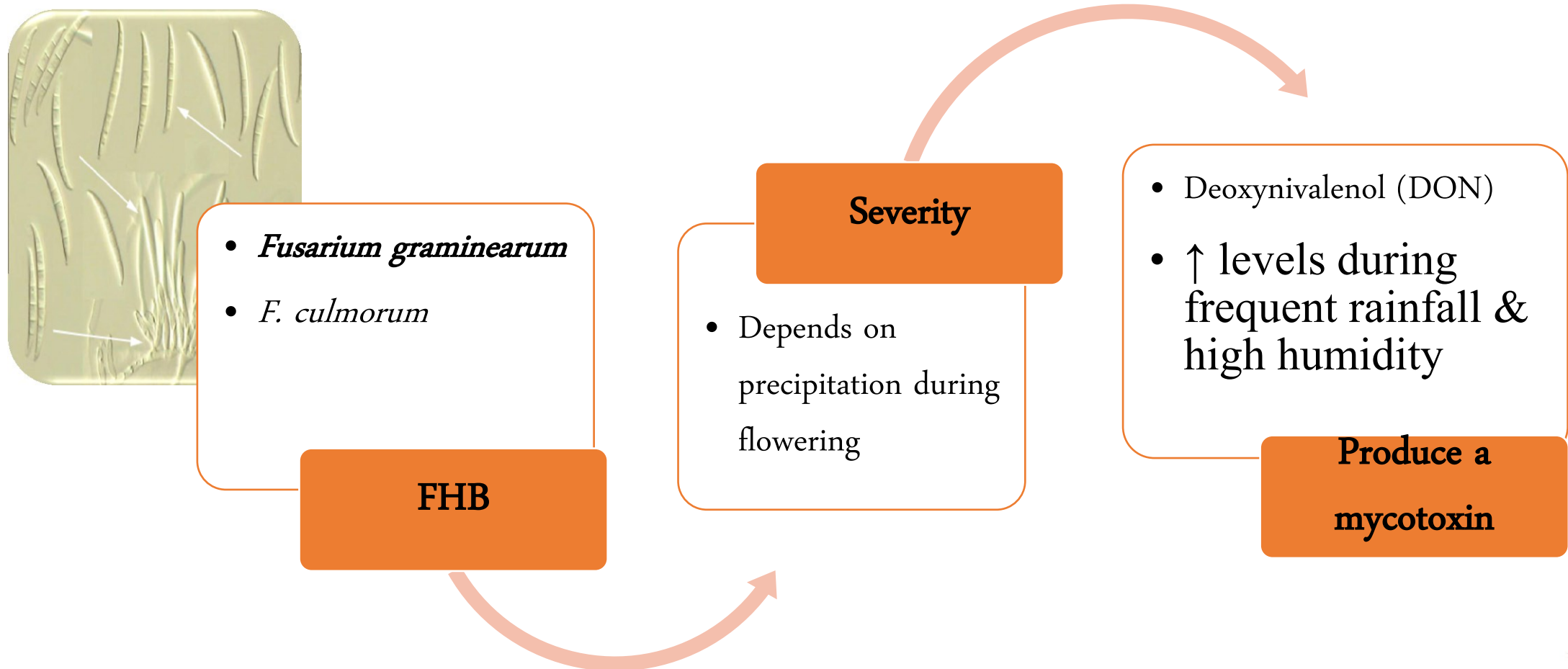


In the **USA** it is **causing economic losses** to the agricultural production and marketing system



Rising cost and risks for growers, inducing them to use more costly management practices or shifting to other crops

FHB & *Fusarium graminearum*



Quantitative trait loci (QTL)-FhBb1 governs resistance towards FHB

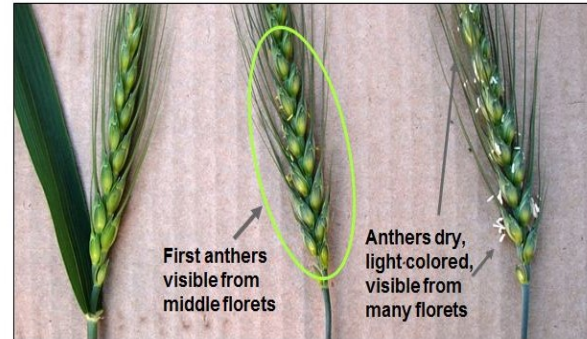
Factors influence germination of wheat & *Fusarium*

DON production



High moisture or relative humidity (>90%) and duration

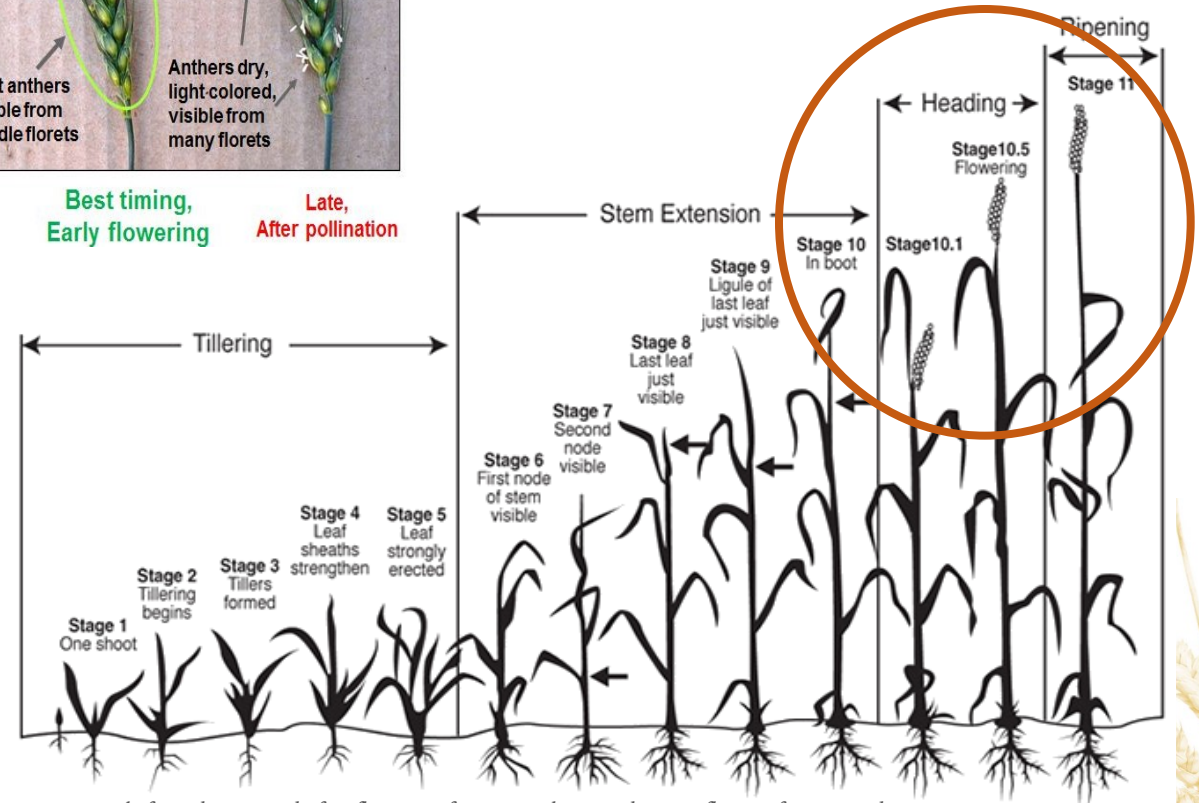
Warm temperatures (5-30 °C/59-86 °F)



Early,
Not yet flowering

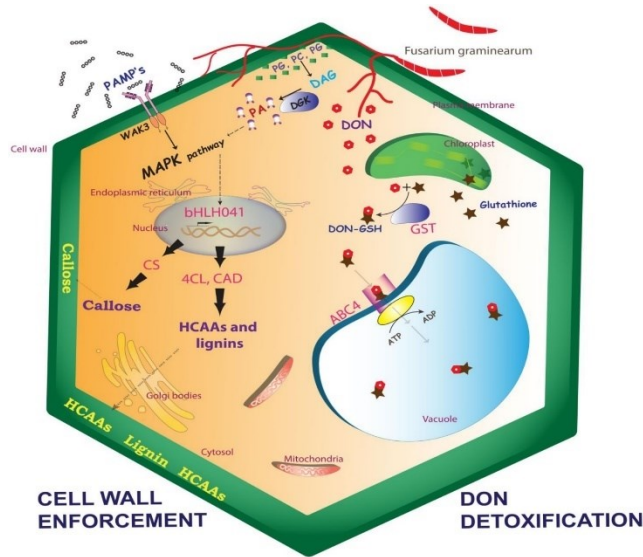
Best timing,
Early flowering

Late,
After pollination



These conditions present before, during, and after flowering favor inoculum production, floret infection, and colonization of developing grains.

Deoxynivalenol-3-glucoside (D3G)



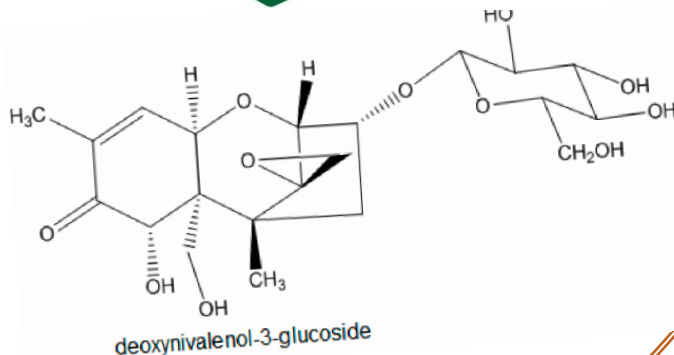
D3G is one the **most common forms of masked DON**

It is **formed as part of a detoxification process in the plant through the glycosylation of DON** and is stored in the plant vacuoles

D3G formation is connected with **glycosyltransferases**

UDP-glycosyltransferase AtUGT73C5 transferred glucose to the OH group at carbon 3 of DON forming D3G in *Arabidopsis* (Poppenger et al. 2003)

It has been **found in wheat lines with low FHB susceptibility: D3G is converted by the quantitative trait locus QTL-Fhb1** (encodes a glycosyltransferase or regulates the expression of such enzyme)



D3G was found in approximately 5% of the food, feed, and unprocessed grains of undefined end-use by European countries. D3G should be measured because it is unknown how this can affect the human and animals health

Objectives

To analyze the DON and D3G content and determine if there is a correlation between the DON and D3G production in samples with variation in susceptibility to FHB grown in Minnesota, USA at two locations and three years of study using liquid chromatography-quadrupole time of flight mass spectrometry (LC-MS)



Wheat samples

WHEAT LINES RANGING FROM MODERATELY SUSCEPTIBLE TO SUSCEPTIBLE TO FHB

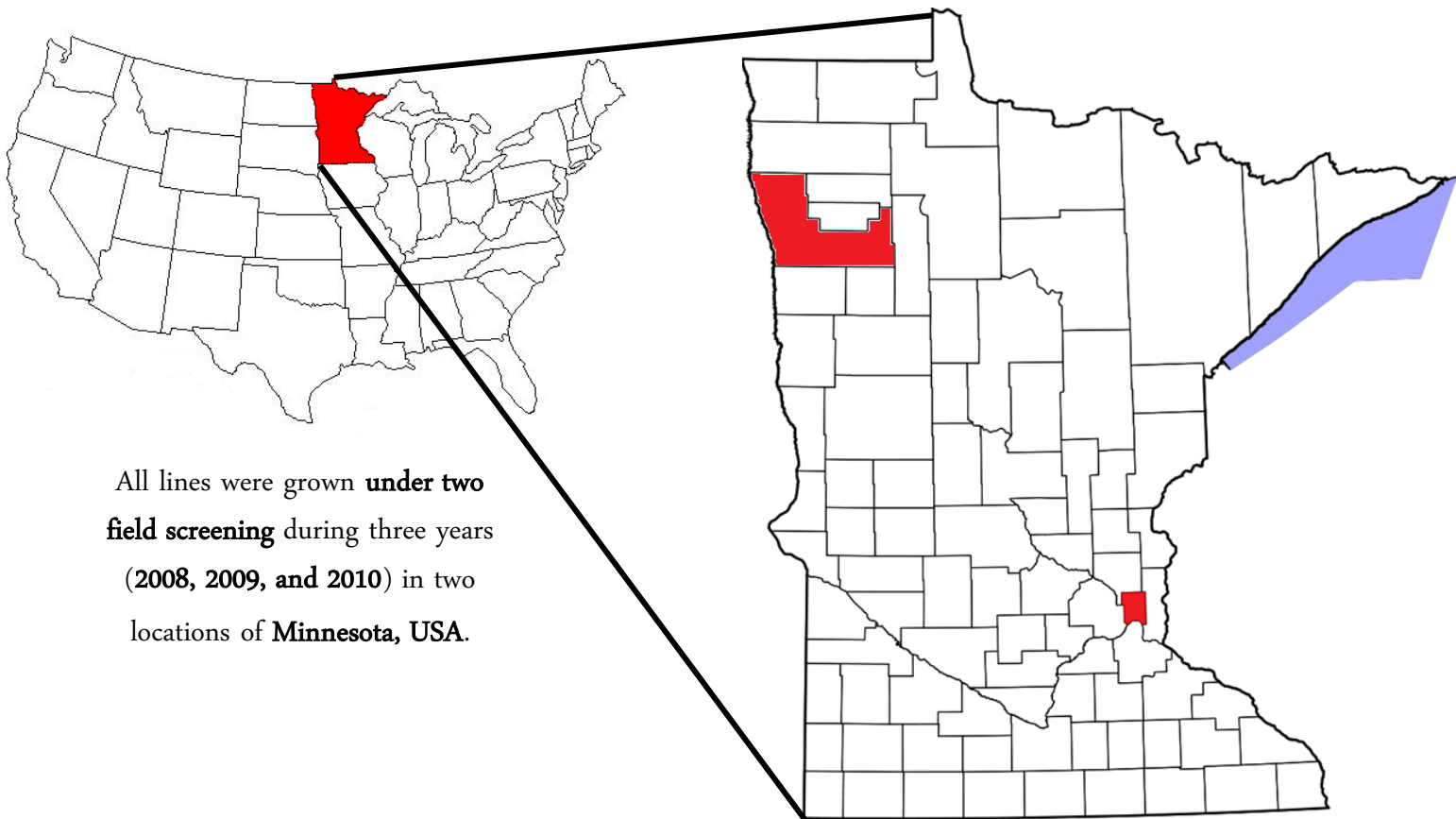
SAMPLES COLLECTED WHEN LATEST MATURING LINES WERE AT HARVEST RIPENESS (14% OR LESS MOISTURE CONTENT)

EXPERIMENTAL SPRING WHEAT LINES (WHEAT BREEDING PROGRAM) RANGING FROM 1ST YEAR TO 3RD YEAR FIELD TRIAL LINES



Growing locations

St. Paul (44.9441° N, 93.0852° W)



All lines were grown **under two field screening** during three years (2008, 2009, and 2010) in two locations of **Minnesota, USA**.

Crookson (47.7742° N, 96.6081° W)

Year	Weather conditions
2008	During Planting: cool and wet wet Growing conditions: hot and slight dry. Adequate soil slight dry. Adequate soil moisture moisture
2009	Growing conditions: Cooler than average with adequate than average with adequate precipitation precipitation
2010	Growing conditions: Cooler growing temperatures and growing temperatures and adequate precipitation adequate precipitation

Fusarium graminearum inoculation

St. Paul (44.9441° N, 93.0852° W)



Backpack sprayer: control timing and inoculum dose

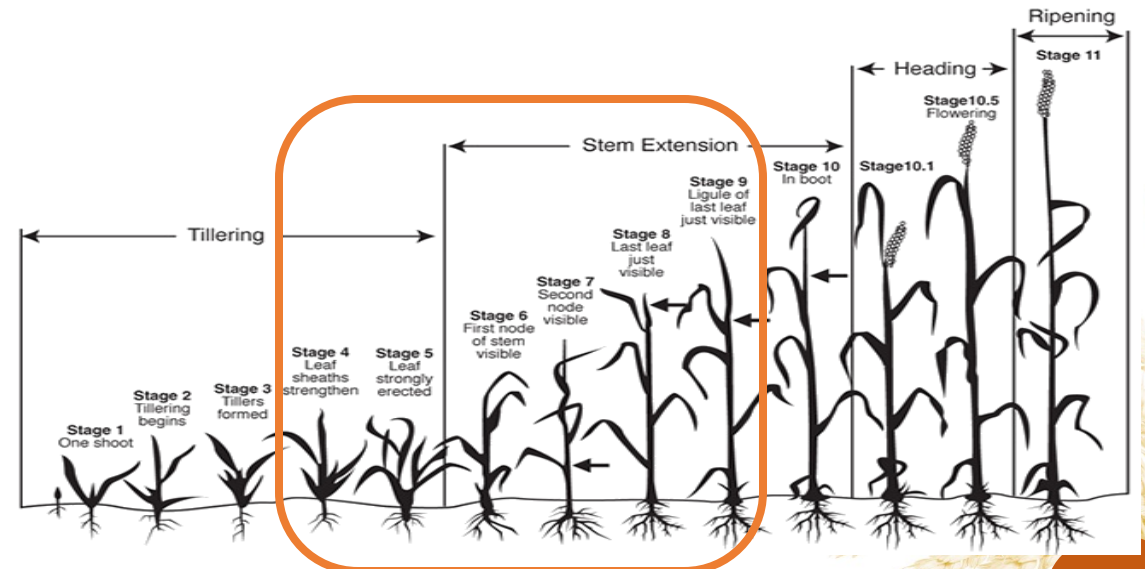
- ❖ *F. graminearum* macrodina applied at the rate of 60 mL
- ❖ 100,000 conidia/mL per 2.4 m row at anthesis and 3-4 days later



Both nurseries were misted periodically overnight to maintain high humidity environments

Crookson (47.7742° N, 96.6081° W)

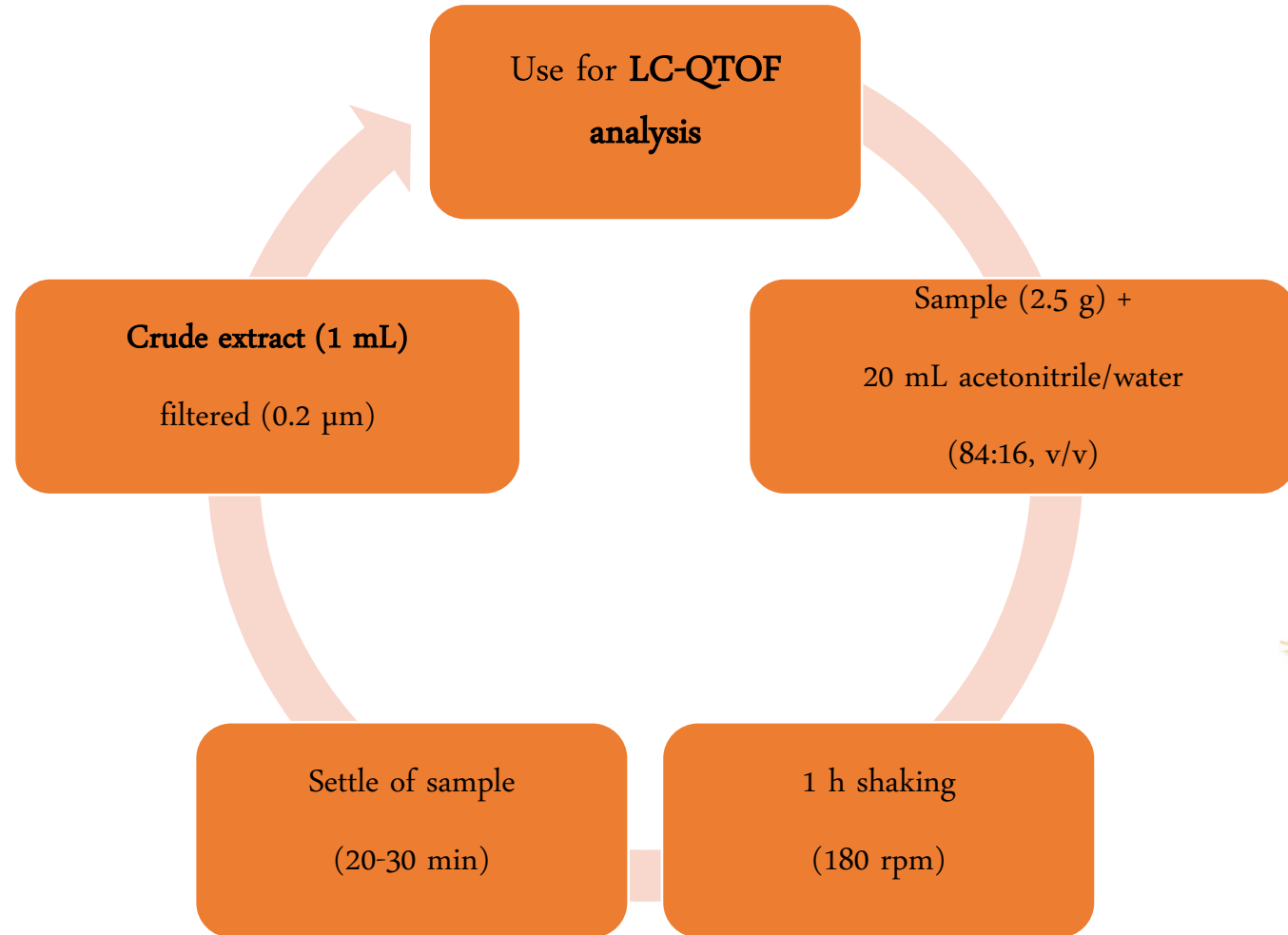
- ❖ Grain spawn inoculum was spread at the rate of 56 kg/ha (jointing stage). Second application one week later.
- ❖ Grain-spawn inoculum method mimics more closely what happens in nature



Sample preparation



0.8 mm screen



DON and D3G analysis with LC-QTOF



1200 series HPLC system (Agilent Technologies)

Eclipse Plus C18 column (2.1 x 100 mm, 1.8-Micron, Agilent Technologies)

Volume injection: 5 μL

LC conditions:

Column temperature: 40 °C

Solvent system:

0.1 % formic acid/water (Solvent A)

0.1% formic acid/acetonitrile (Solvent B)

Gradient:

Purge: 100% A, flow rate: 4 mL/min, 15 s

Flow rate: 0.4 mL/min

97% A and 3% B, 0.75 min

Linear increase to 100% B, 4 min

Hold time 100% B, 6 min

Re-equilibration at 97% A, 10 min

Washout with 100% A, 2 min

DON and D3G analysis with LC-QTOF



Mass spectrometer quadrupole time of flight (Agilent 1290 series LC/MS)

ESI interface in positive ionization mode at 300 °C

Settings:

Gas flow: 7 L/min

Nebulizer gas: 30psig

225 sheath gas temperature and 12 of sheath gas flow

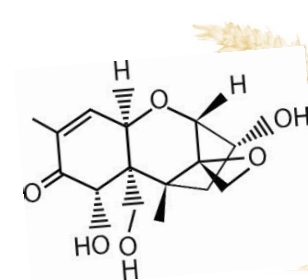
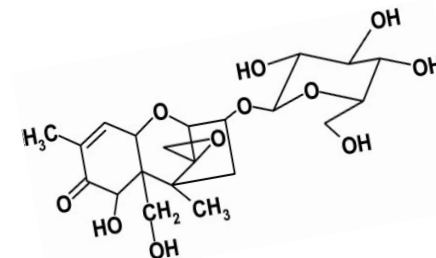
Mode MSI parameters

Minimal range(m/z): 100

Maximum range(m/z): 1700

Scan range: 2 spectra/s

Data analysis: MassHunter Qualitative analysis B.05.00
program (Agilent Technologies)



Statistical analysis

ANOVA was performed individually for three year data in SAS (V 9.2, SAS Institute, Inc., Cary, NC, USA).

The GLM was used for ANOVA in which wheat line and location were considered as fixed effects.

The main effects of wheat line and location and their interaction were tested for significance using the residual error terms.

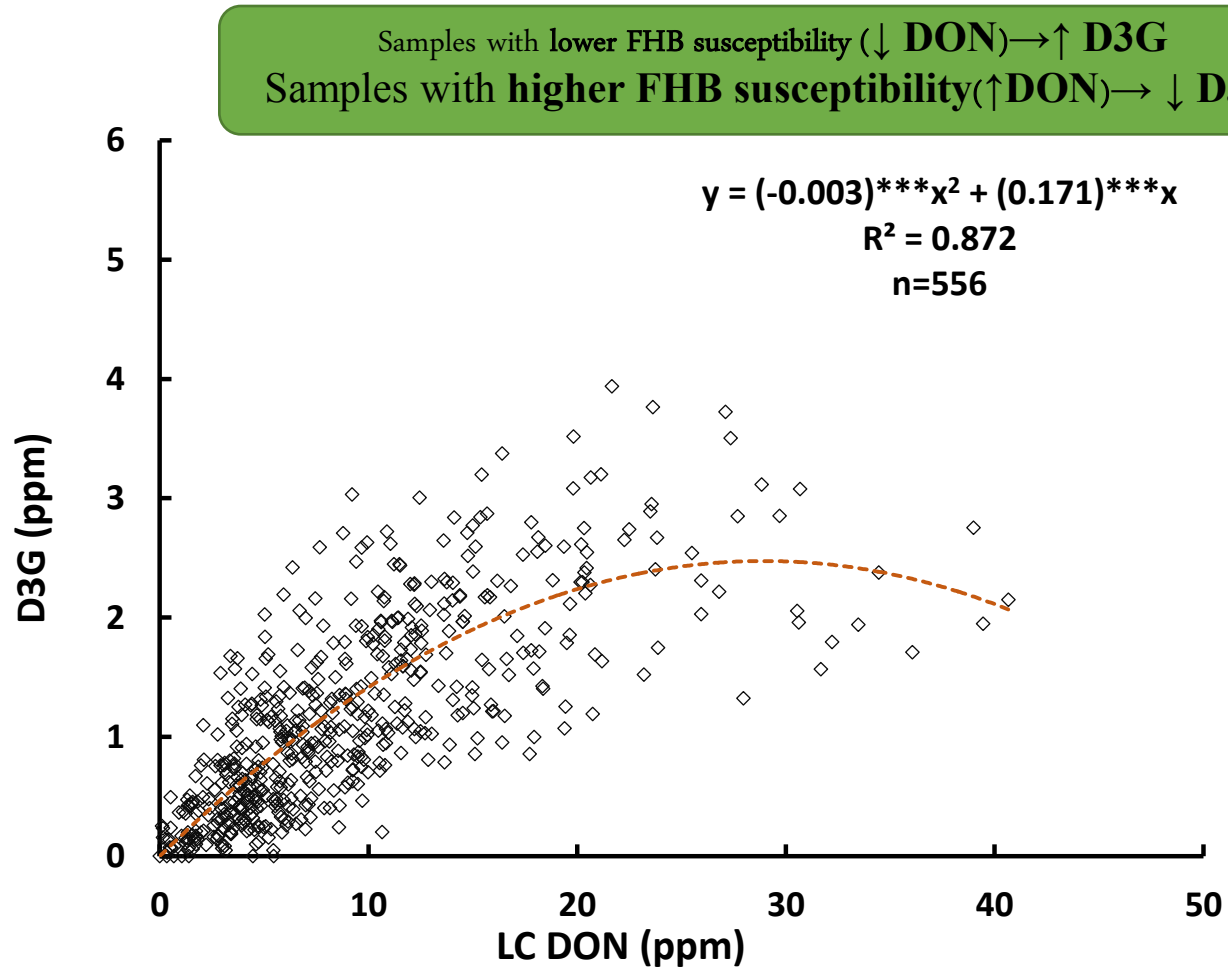
Correlation and regression was performed using “CORR” and “GLM” procedure in SAS.



Results and discussions



Relationship between DON and D3G



*** significantly different from 1 at $p < 0.001$

There was a correlation between DON and D3G (**second order curve**)

D3G content rose as DON content increased in samples with DON content: 0-30 ppm

But at **higher DON** concentration, a decrease in the D3G content was observed

The **type of inoculum** used between locations **and inoculation at slightly different growth stages** could be causing this behavior

Effect of the line, location and the

DON: 0.1 to 33.9 ppm

D3G: 0.1 to 1.9 ppm

DON: 0.0 to 23.6 ppm

D3G: 0.0 to 3.0 ppm

DON: 0.2 to 17.7 ppm

D3G: 0.0 to 2.2 ppm

Lowest mycotoxin contents

Year

Location

Range

DON

D3G

2008

Crookston

Min (n=22)

0.1

0.3

Max (n=22)

24.2

1.8

Average (n=22)

5.7

1.1

St. Paul

Min (n=22)

0.7

0.1

Max (n=22)

39.5

1.9

Average (n=22)

11.1

0.9

MN

Min (n=44)

0.1

0.1

Max (n=44)

39.5

1.9

Average (n=44)

8.4

1.0

2009

Crookston

Min (n=35)

0.0

0.0

Max (n=35)

25.7

3.8

Average (n=35)

11.9

2.1

St. Paul

Min (n=35)

0.2

0.0

Max (n=35)

21.0

1.5

Average (n=35)

4.8

0.5

MN

Min (n=70)

0.0

0.0

Max (n=70)

25.7

3.8

Average (n=70)

8.3

1.3

2010

Crookston

Min (n=88)

1.7

0.4

Max (n=88)

20.2

2.6

Average (n=88)

7.9

1.3

St. Paul

Min (n=90)

0.2

0.0

Max (n=90)

11.5

1.5

Average (n=90)

4.2

0.5

MN

Min (n=178)

0.2

0.0

Max (n=178)

20.2

2.6

Average (n=178)

6.1

0.9

ppm, n=number of lines in each set.

ANOVA table for DON and D3G of wheat samples for 2008-2010

Year	Traits	Source	DF	Sum of Squares	Mean square	F value	Pr > F
2008	DON	Line	21	3369.6	160.5	2.9	0.0095
		Loc	1	418.7	418.7	7.5	0.0122
		Line*Loc	21	1169.0	55.7	3.5	0.015
		Error	12	191.7	16.0		
	D3G	Line	21	12.7	0.60	3.8	0.0016
		Loc	1	0.9	0.88	5.6	0.0275
		Line*Loc	21	3.3	0.16	4.7	0.004
		Error	12	0.4	0.03		
2009	DON	Line	34	5314.4	156.3	5.3	< 0.001
		Loc	1	1013.4	1013.4	34.3	< 0.001
		Line*Loc	34	1004.5	29.5	1.3	0.177
		Error	72	1639.7	22.8		
	D3G	Line	34	27.9	0.82	2.5	0.0048
		Loc	1	49.3	49.28	149.0	< 0.0001
		Line*Loc	34	11.2	0.33	2.6	0.000
		Error	72	9.0	0.13		
2010	DON	Line	89	3479.0	39.1	8.0	< 0.0001
		Loc	1	650.1	650.1	132.5	< 0.0001
		Line*Loc	87	426.8	4.9	1.0	0.497
		Error	88	419.0	4.8		
	D3G	Line	89	48.1	0.54	4.0	< 0.0001
		Loc	1	33.1	33.09	247.4	< 0.0001
		Line*Loc	87	11.6	0.13	1.0	0.497
		Error	88	11.8	0.13		

DON and D3G content not statistically related to the main effects (Line*Loc)

The effects of Line and Loc on DON were statistically significant → growing location is the main influence of DON content among samples

The effects of Line and Loc were significantly related to the DON and D3G content in the samples, but there was not interaction between factors → genetic and environmental conditions play an important role in the DON and D3G production

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DON:

DON **dependent on the growing conditions** in a particular season, is **affected by the wheat line**. However, **the main influence is the location** where the wheat line is planted, **which also showed highest influence on the D3G content**

D3G:

The **D3G production will depend on the tolerance/susceptibility level** of the wheat line **to FHB** and its **response to environmental conditions**

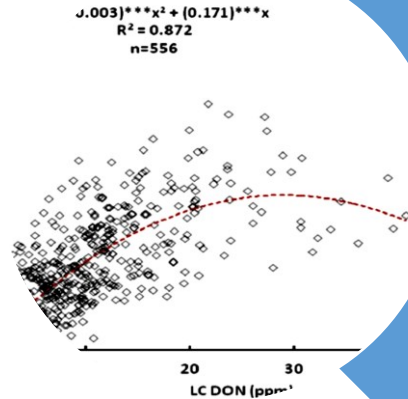
The results on **D3G content could help to increase the data about its occurrence in hard red spring wheat** in the USA, in terms of **food safety** because this masked mycotoxin **might be converted back to DON**

Pearson and Spearman's correlation coefficients between DON and D3G of two localities of Minnesota for 2008-2010

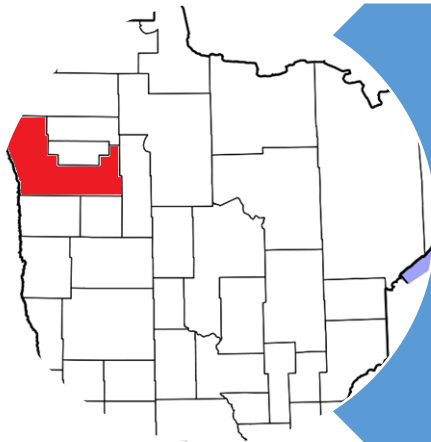
Year	Crk DON	Stp DON	Crk D3G	Stp D3G
2008	Pearson correlation			
	-	0.59 **	0.56 **	0.56 **
	0.51 *	-	0.47 *	0.90 ***
	0.68 ***	0.46 *	-	0.59 **
	0.58 **	0.87 ***	0.51 *	-
2009	Spearman correlation			
	Pearson correlation			
	-	0.52 **	0.66 ***	0.50 **
	0.45 **	-	0.24 NS	0.75 ***
	0.56 ***	0.44 **	-	0.41 *
2010	Spearman correlation			
	Pearson correlation			
	-	0.63 ***	0.69 ***	0.41 ***
	0.57 ***	-	0.51 ***	0.67 ***
	0.55 ***	0.45 ***	-	0.48 ***
2010	0.34 **	0.61 ***	0.44 ***	-
	Spearman correlation			

Crk: Crookston, Stp: Saint Paul, NS: no significant. *, **, ***: means correlation coefficient is significant at $p < 0.05$, 0.01 , 0.001 .

Conclusions



THE RELATIONSHIP BETWEEN DON AND D3G FIT A SECOND ORDER CURVE, INDICATING THAT THE TOLERANCE OF THE WHEAT LINES TO THE FUSARIUM INFECTION IS RELATED TO THE ABILITY OF THE WHEAT LINE TO CONVERT THE DON TO D3G DURING THE DETOXIFICATION PROCESS



THE MOST IMPORTANT FACTOR AFFECTING THE DON AND D3G FORMATION IS LOCALITY, WHICH MAY BE DUE TO DIFFERENCES IN GENE EXPRESSION OF THE WHEAT LINE IN DIFFERENT ENVIRONMENTAL CONDITIONS AND ITS RESPONSE TO DIFFERENT INOCULUM AND DEVELOPMENT STAGES OF THE WHEAT DURING THE INOCULATION PROCESS

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Thank you for listening!

