



Mycotoxins and Health

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Wheat Quality Laboratory



NDSU Wheat
Quality Lab

Project Leader
Dr. Senay Simsek

Survey: Regional
Quality Report

Export Cargo
Survey

Nursery Samples:
Breeding Programs

Research Projects



Including Field
Plots

Established 1907

STUDENT FOCUSED • LAND GRANT • RESEARCH

NDSU

My Research Team

Graduate Students:

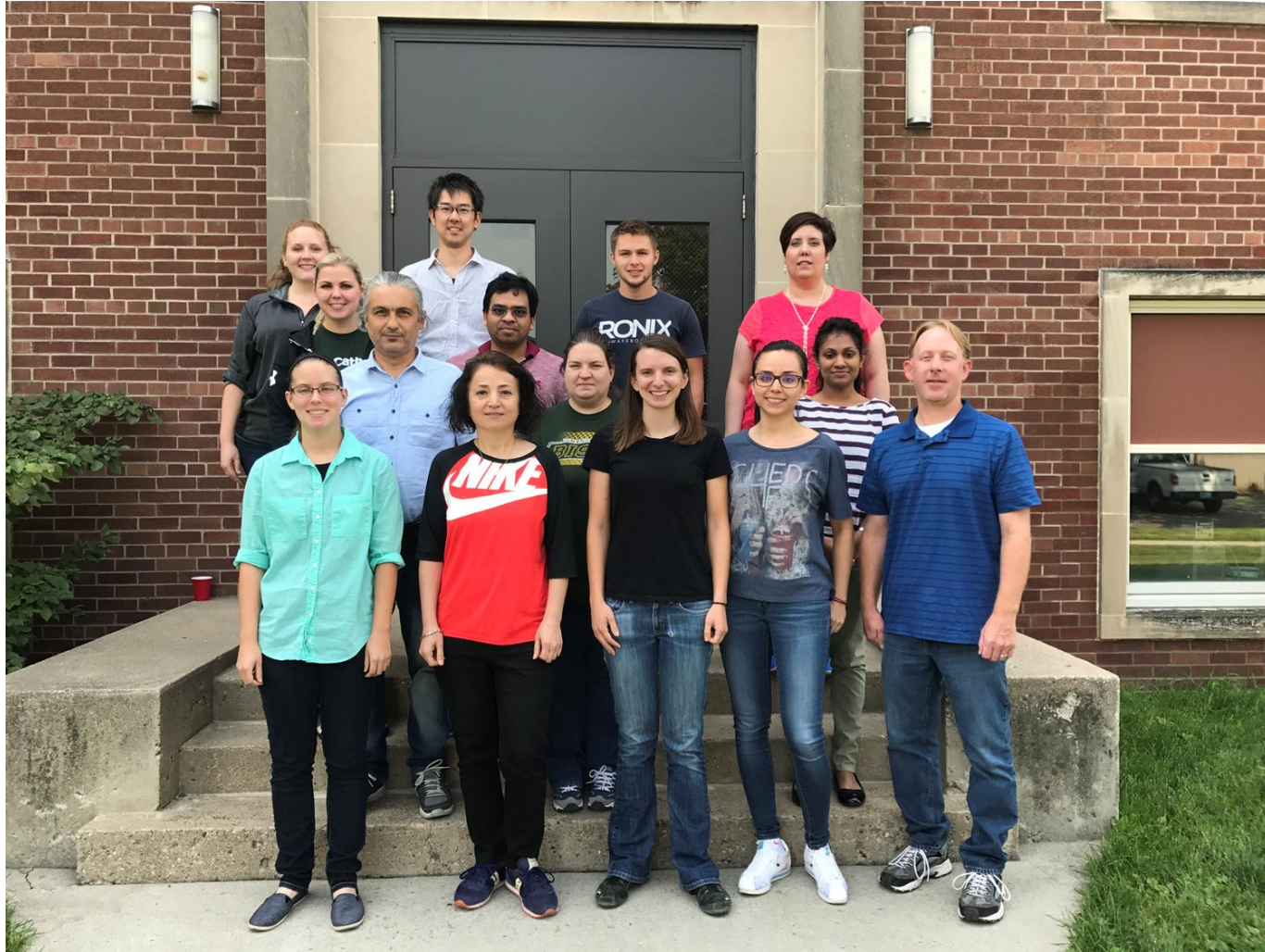
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Md Mahfuzur Rahman (MS)
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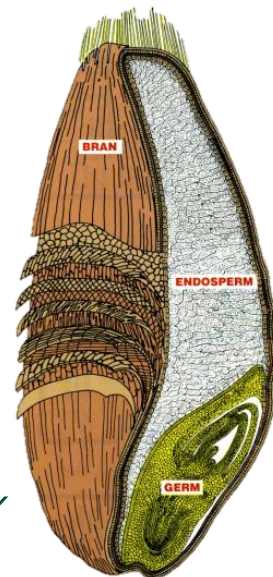
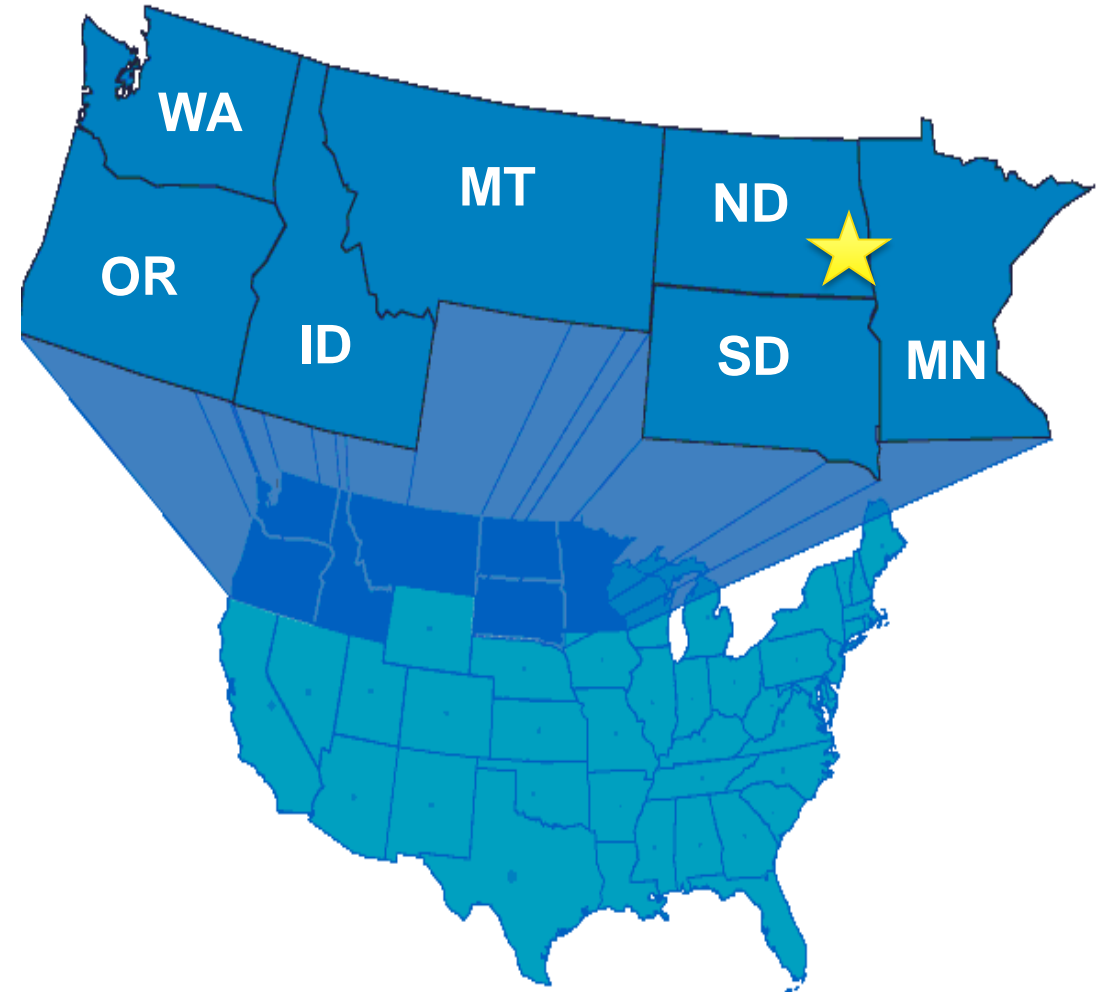


FUSARIUM HEAD BLIGHT

Hard Red Spring Wheat

Specialty wheat grown in the Northern Plains of United States

- Highest protein content
- Hard endosperm
- Red bran
- Strong gluten
- High water absorption



NORTH DAKOTA
STATE UNIVERSITY

Mold

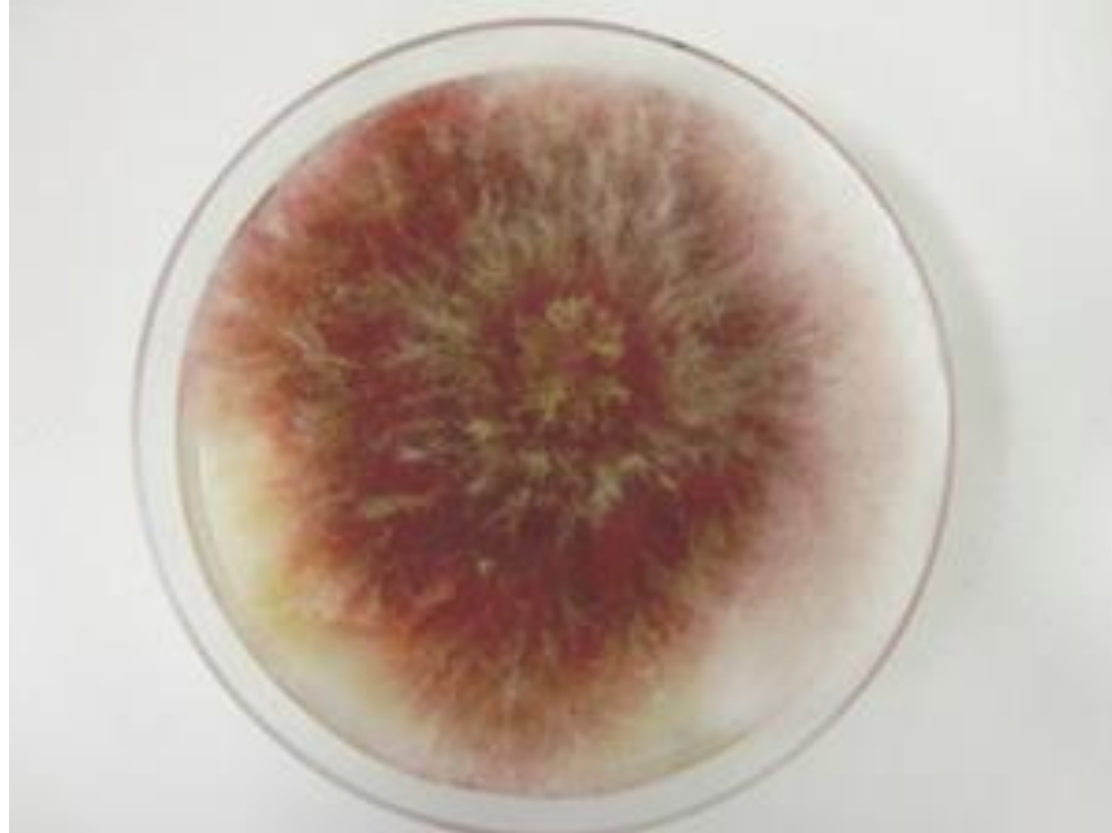


Figure 1. *Fusarium graminearum* mycelium growth on artificial growth media.

Nebraska Extension. 2012. Major *Fusarium* diseases on corn, wheat, and soybeans in Nebraska. <http://extensionpublications.unl.edu/assets/html/g2181/build/g2181.htm> (accessed 2017 December 17).

Important Definitions



Toxin

Toxicology

Infection vs intoxication

***FUSARIUM GRAMINEARUM* INFECTIONS IN GRAIN**



Wheat infected with *Fusarium graminearum*.

Wisconsin Field Crops Pathology. *Fusarium* head blight (scab) of wheat.
<https://fyi.uwex.edu/fieldcroppathology/fusarium-head-blight-scab-of-wheat/> (accessed 2017 December 17).

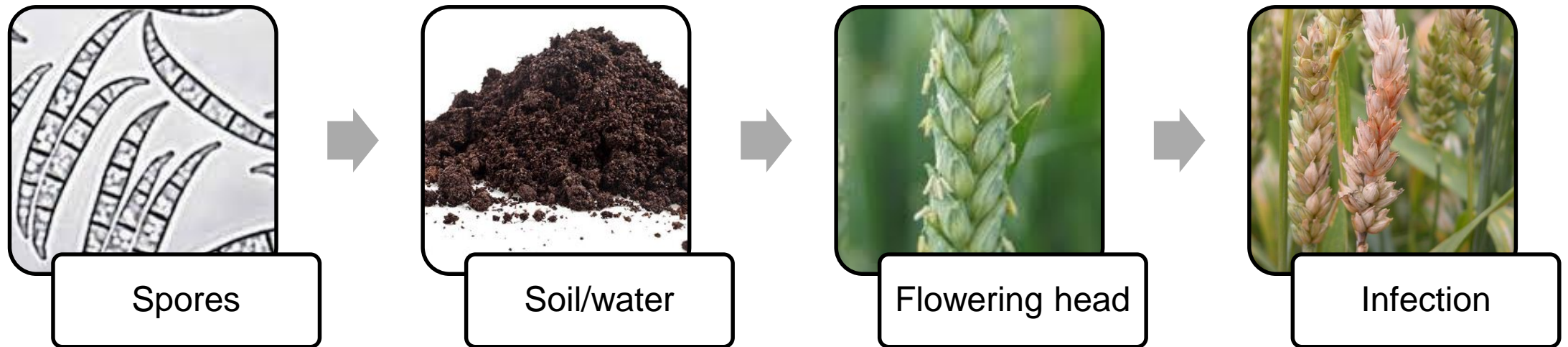


Fusarium graminearum mycelium growing on germinating wheat.

Jin, Z., Zhou, B., Gillespie, J., Gross, T., Barr, J., Simsek, S., Brueggeman, R., and Schwarz, P. 2018. Production of deoxynivalenol (DON) and DON-3-glucoside during the malting of *Fusarium* infected hard red spring wheat. Food Control. 85:6-10.

Wheat infected with *Fusarium graminearum*

Fusarium Head Blight (FHB) Cycle



Disease cycle of Fusarium head blight.

Ohio State University Extension. 2017. *Fusarium* head blight or head scab on wheat, barley, and other small grains. <https://ohioline.osu.edu/factsheet/plpath-cer-06> (accessed 2017 December 17).

Fusarium graminearum Growth

Crops

- Wheat
- Barley
- Corn
- Rye
- Oat

Weather

- Warm
- Moist
- Humid

Farming style

- Tilling
- Crop rotation
- Fungicide

Production of DON by *F. graminearum*

Highly variable

Temperature, water activity, pH, nutrient availability, etc.

Optimum conditions

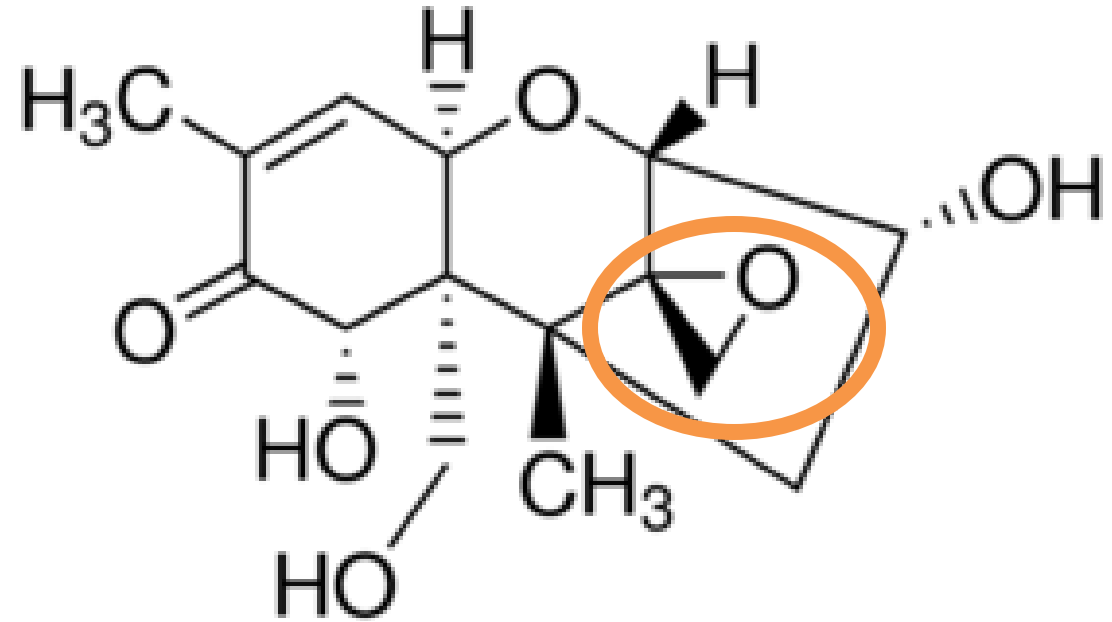
pH 4.91 and 23.75 °C

Inducer of DON production

Expression of the *Tri5* gene

Deoxynivalenol (DON)

- Mycotoxin produced by *Fusarium graminearum*
- Vomitoxin
- 12,13-epoxy trichothecene
- Water soluble toxin



Chemical structure of deoxynivalenol.

Sigma Aldrich. 2017. Deoxynivalenol.
<https://www.sigmaaldrich.com/catalog/product/sigma/d0156?lang=en®ion=US> (accessed 2017 December 17).

FDA Limits on DON in Food and Feed

Table 3. FDA limits on DON in food and feed.

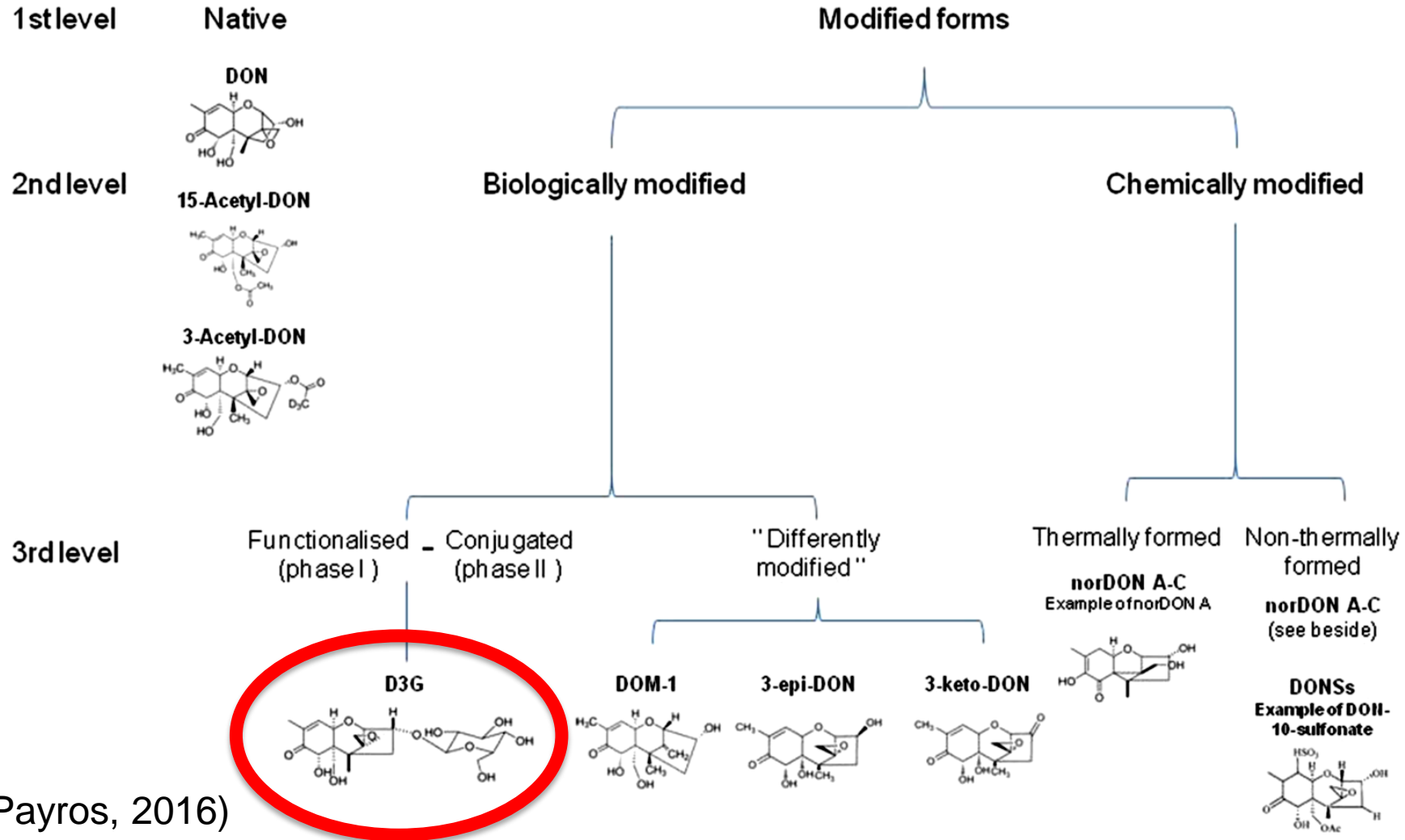
Product	DON limit (ppm)
Wheat products for humans	1
Feed for pigs (<20 % of diet)	5
Grain for chicken (<50 % of diet)	10
Grain for cattle (<50 % of diet)	10
Distillers grain and gluten feed (Cattle >4 months old)	30
Grain for all other animals (<40 % of diet)	5
Raw grain intended for processing	No limit

Modification of Mycotoxins

- Biological and chemical changes can modify mycotoxin structure
- “Modified mycotoxins”
 - Different forms of mycotoxins
- “Masked mycotoxins”
 - Only toxins conjugated by plants

(Payros, 2016)

Modification Mechanism



Bound Mycotoxins

Deoxynivalenol-3-glucoside (D3G)

- **3- β -D-glucopyranosyl-4-deoxynivalenol**

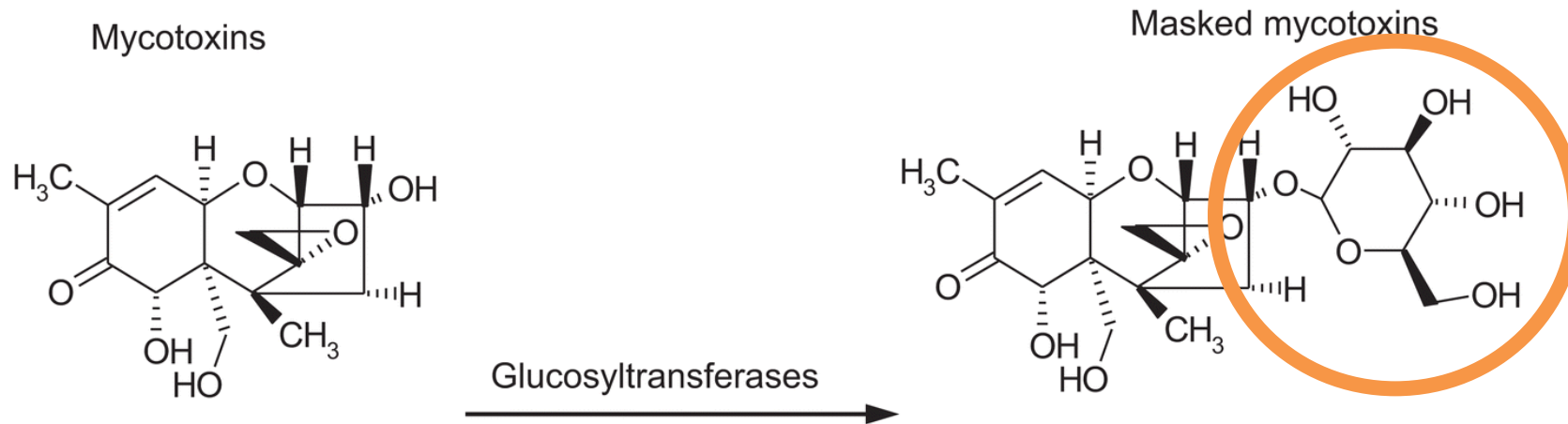
Produced by the plant as a detoxification method

Samples with DON may have DON-3-glucoside

D3G/DON \approx 10-30%.

DON-3-glucoside (D3G)

- Phase II metabolism product
- Less toxic than DON
- Stored in plant vacuoles
- Masked mycotoxin



Reaction mechanism of the formation of deoxynivalenol-3-glucoside from deoxynivalenol.

Agricultural Costs of *Fusarium graminearum* and DON

Decreases

- Yield, quality, and overall grade

Financial losses

- Value of yield lost: \$1 billion
- Testing at elevators: \$21 million
- Added costs at mills: \$8 million

DON, D3G AND HEALTH

Illness from Deoxynivalenol

Acute
exposure

Diarrhea

Vomiting

High doses

Circulatory
shock

Reduced
cardiac
output

Death

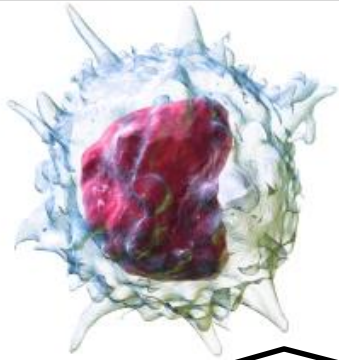
Chronic
exposure

Anorexia

Reduced
weight gain

Altered
nutritional
efficiency

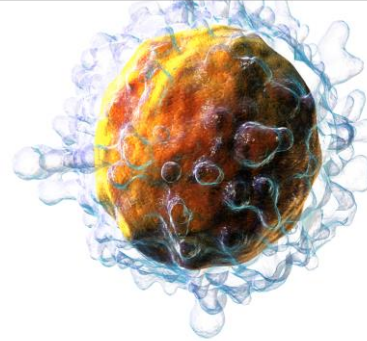
Cellular Targets Of DON



Monocyte



Macrophage



T-lymphocyte

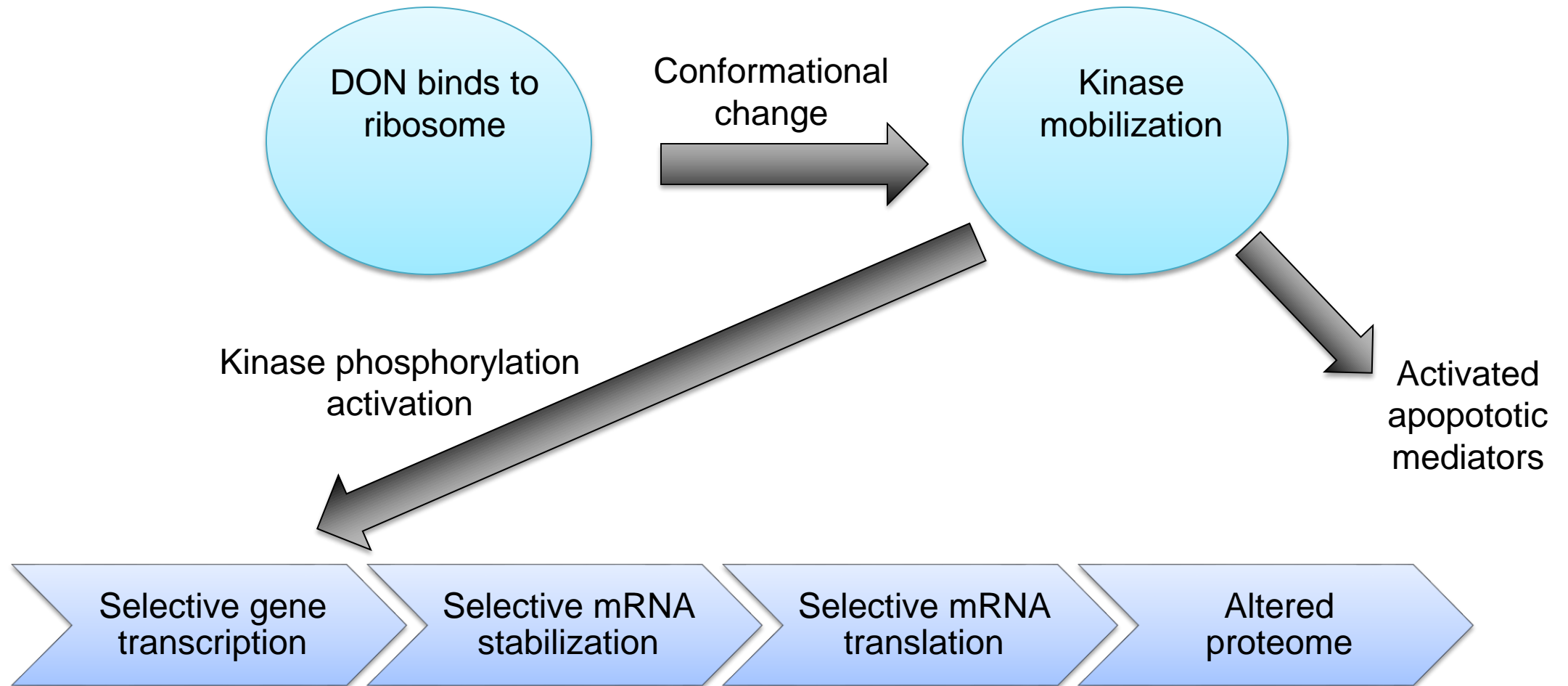


B-lymphocyte

Cellular targets of DON.

Selfhacked 2018. Danger of high or low monocytes. <https://selfhacked.com/blog/monocytes/> (accessed 2018 January 18).

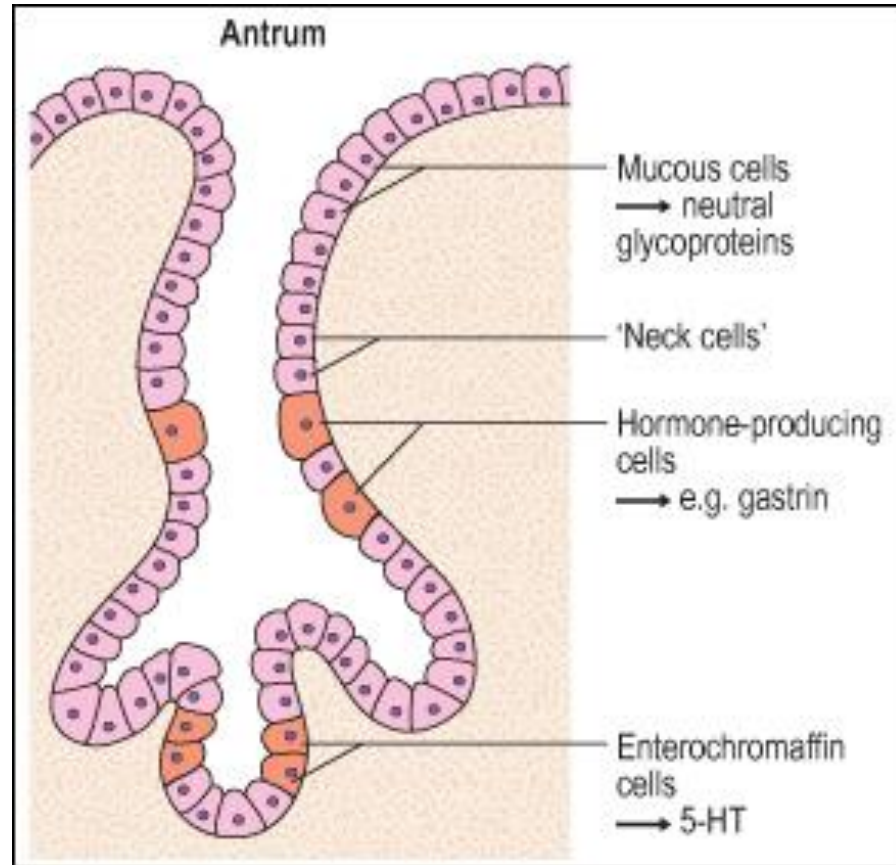
Ribotoxic Stress Response



Ribotoxic stress response overview.

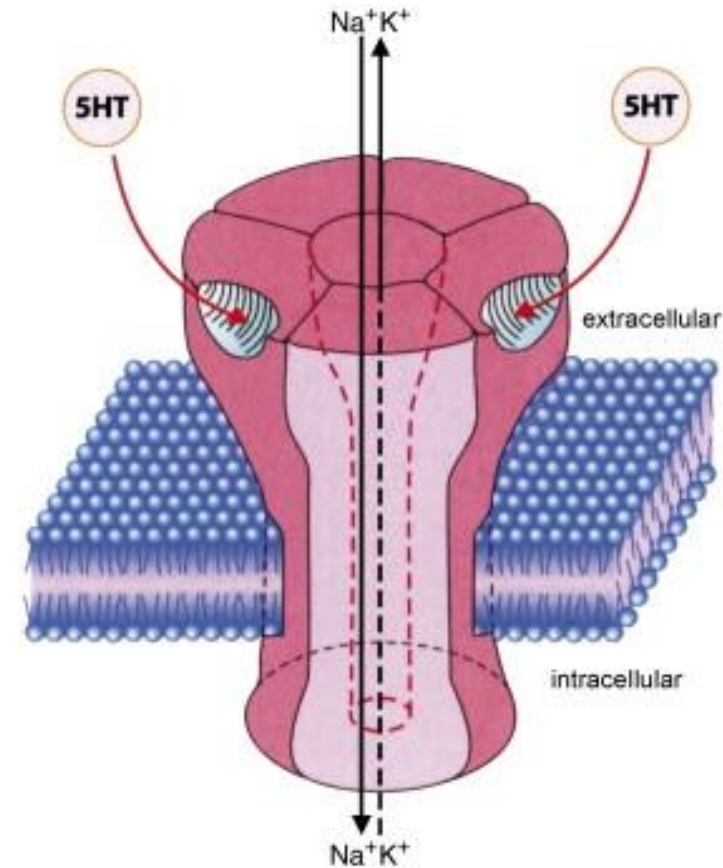
Pestka, J. J. 2010. Deoxynivalenol: Mechanisms of action, human exposure, and toxicological relevance. Arch. Toxicol. 84:663-679.

Reduced Food Intake



Enterochromaffin cells.

The Rise of Caliphate. 2010. L5-Patho peptic ulcer disease. <https://ak47boyz90.wordpress.com/2010/09/02/l5-patho-peptic-ulcer-disease/> (accessed 2017 December 21).



Serotonin receptor.

Faerber, L., Drechsler, S., Ladenburger, S., Gschaidmeier, H., and Fischer, W. 2007. The neuronal 5-HT₃ receptor network after 20 years of research – Evolving concepts in management of pain and inflammation. *Eur. J. Pharmacology*. 560(1):1-8.

Altered Nutrient Uptake



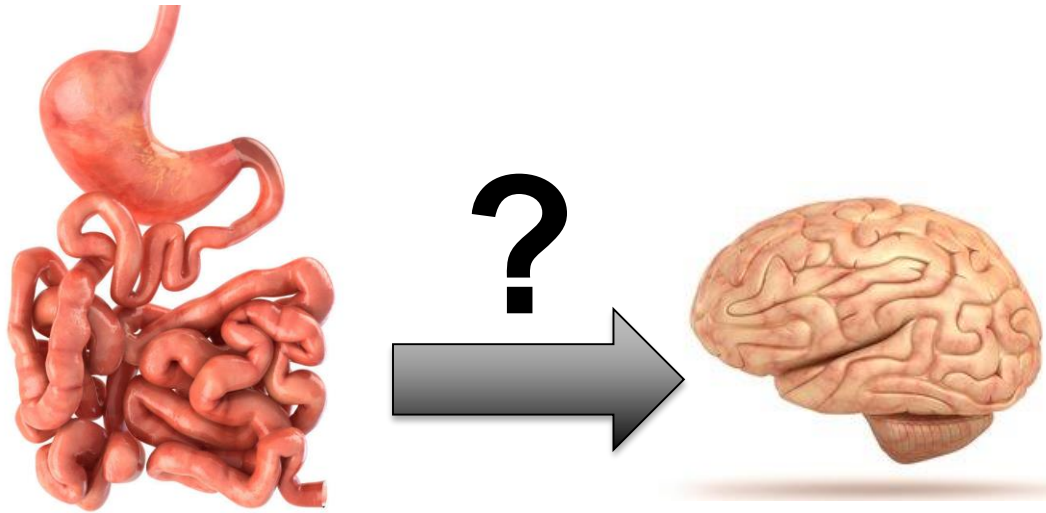
Shortened villi

Enterocyte lysis

Glucose/galactose transporter inhibition

Inhibition of fructose, serine, and glucose transporters

Vomiting



Vomiting due to ingestion of
DON.

Chen, L., Peng, Z., Nussler, A. K., Liu, L., and Yang, W. 2017. Current and prospective sights in mechanism of deoxynivalenol-induced emesis for future scientific study and clinical treatment. J. Appl. Toxicol. 37:784-791.

Norepinephrine ↑



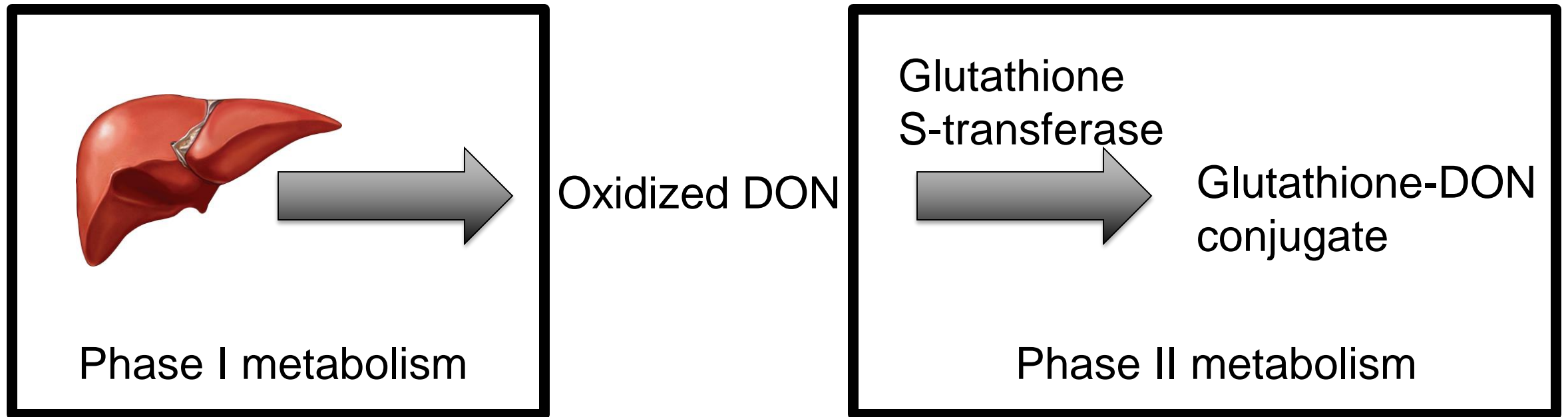
Dopamine ↓



Serotonin ↑



Detoxification of DON



Deoxynivalenol detoxification.

Sobrova, P., Adam, V., Vasatkova, A., Beklova, M., Zeman, L., and Kizek, R. 2010. Deoxynivalenol and its toxicity. Interdiscip. Toxicol. 3:94-99.

Species Susceptibility to DON



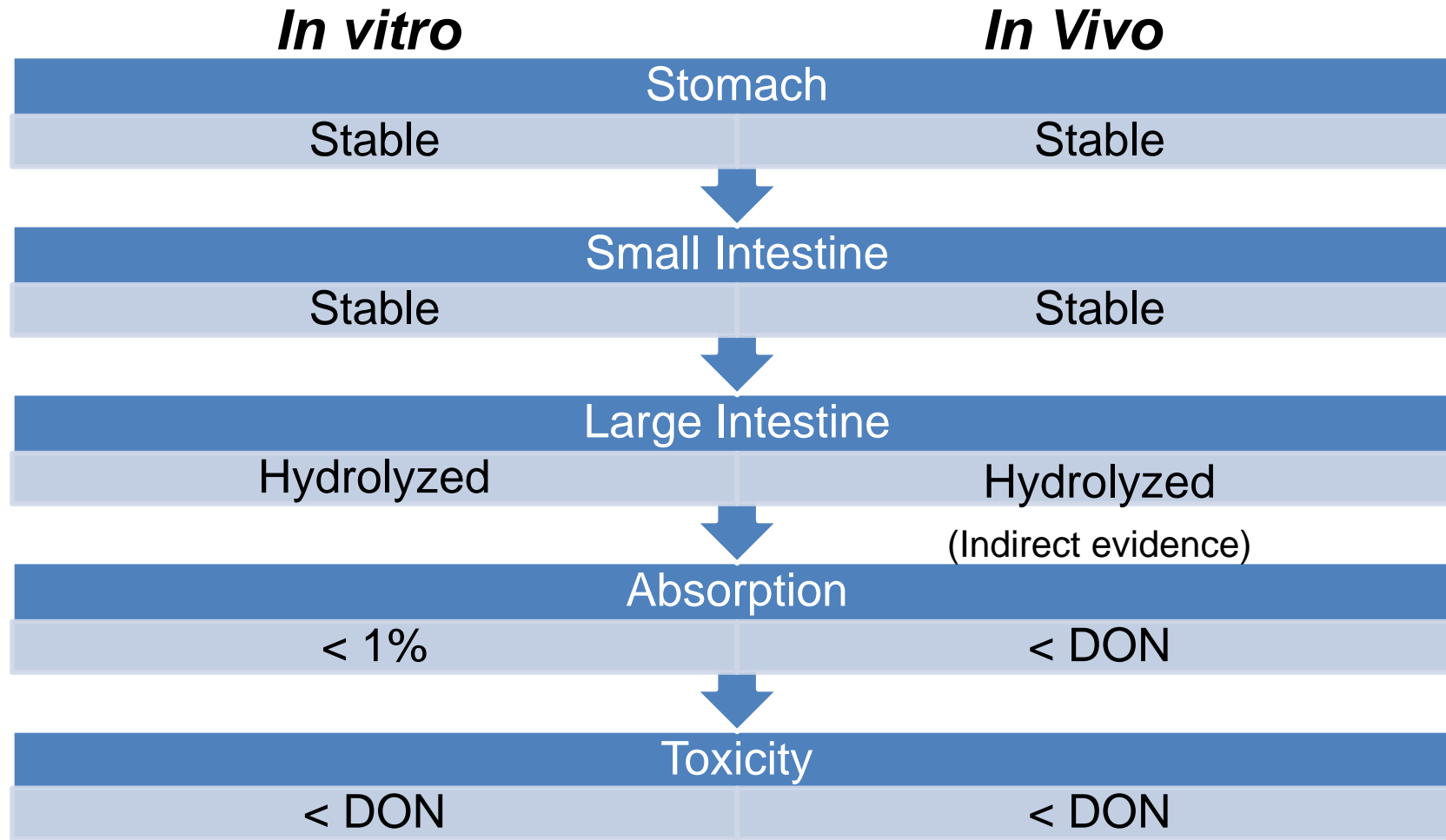
Table 2. Species susceptibility to deoxynivalenol.

Animal	LD ₅₀ (mg kg ⁻¹ body weight)	Administration method
Mice	46-78	Oral
1-day old chicken	140	Oral

Digestion of D3G

- Limited studies on
 - Fate of D3G and other masked mycotoxins in the gut
 - Contribution to toxicity

Digestion and Absorption



Toxicity of D3G

- Thought to have limited bioavailability in the gut
 - Due to covalent binding
- D3G cannot sterically bind to ribosomal 60S subunit A-site pocket
 - Target for DON-induced ribosomal toxicity

Gratz 2017

Toxicity of D3G

- D3G
 - Does not induce ribotic stress
 - No activation of JNK and p38 MAP kinasis
 - Does not alter viability and barrier function of intestinal cells
 - Does not induce morphological lesions or pro-inflammatory gene expression in explants

(Payros, 2016)

D3G Hydrolysis by Fecal Microbiota

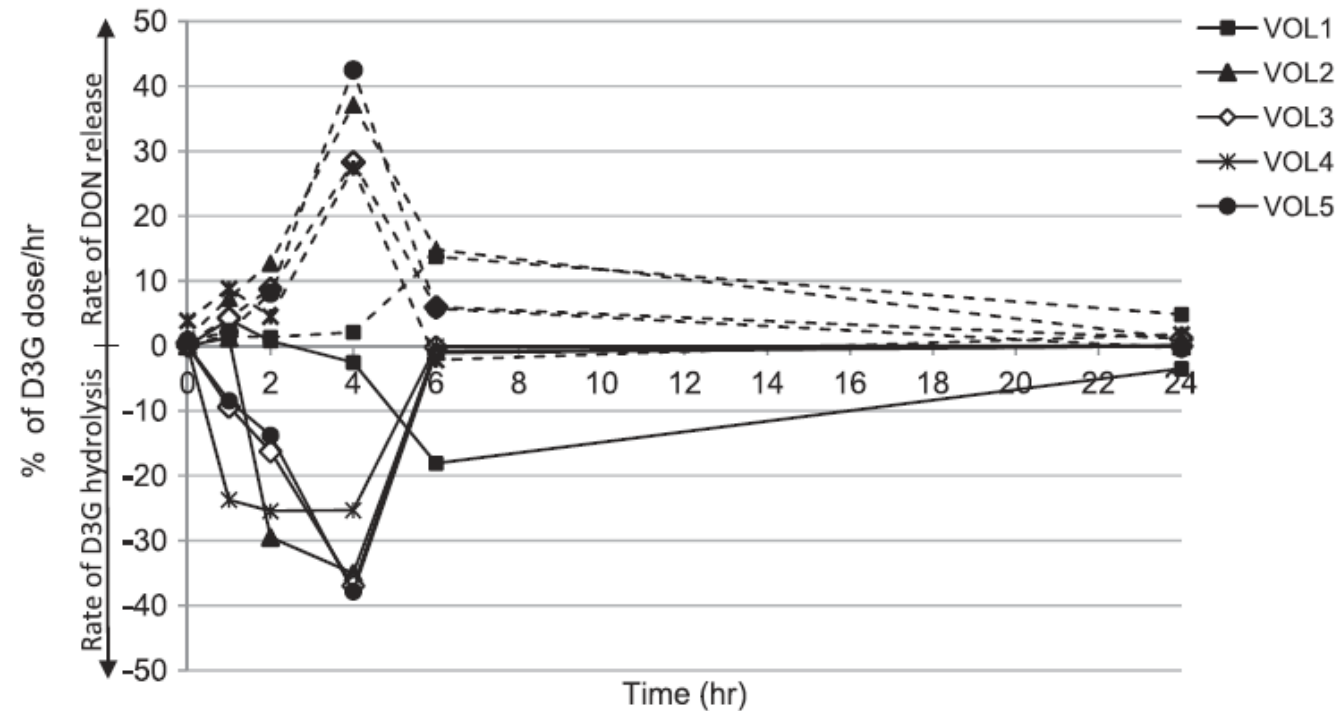


FIG 1 Rate of hydrolysis of D3G and release of DON by the fecal microbiota of 5 volunteers (VOL). Data are presented as D3G hydrolyzed per hour (% of D3G dose [solid lines]) and DON released per hour (% of D3G dose [dashed lines]).

D3G Hydrolysis by Fecal Microbiota

- Fecal microbiota cleave D3G and release DON
- Hydrolysis rates range from 25-38% per hour after 4 hour incubation
- 100% D3G is degraded after 6 hours

Gratz et al., 2013

D3G Metabolism by Human Microbiota

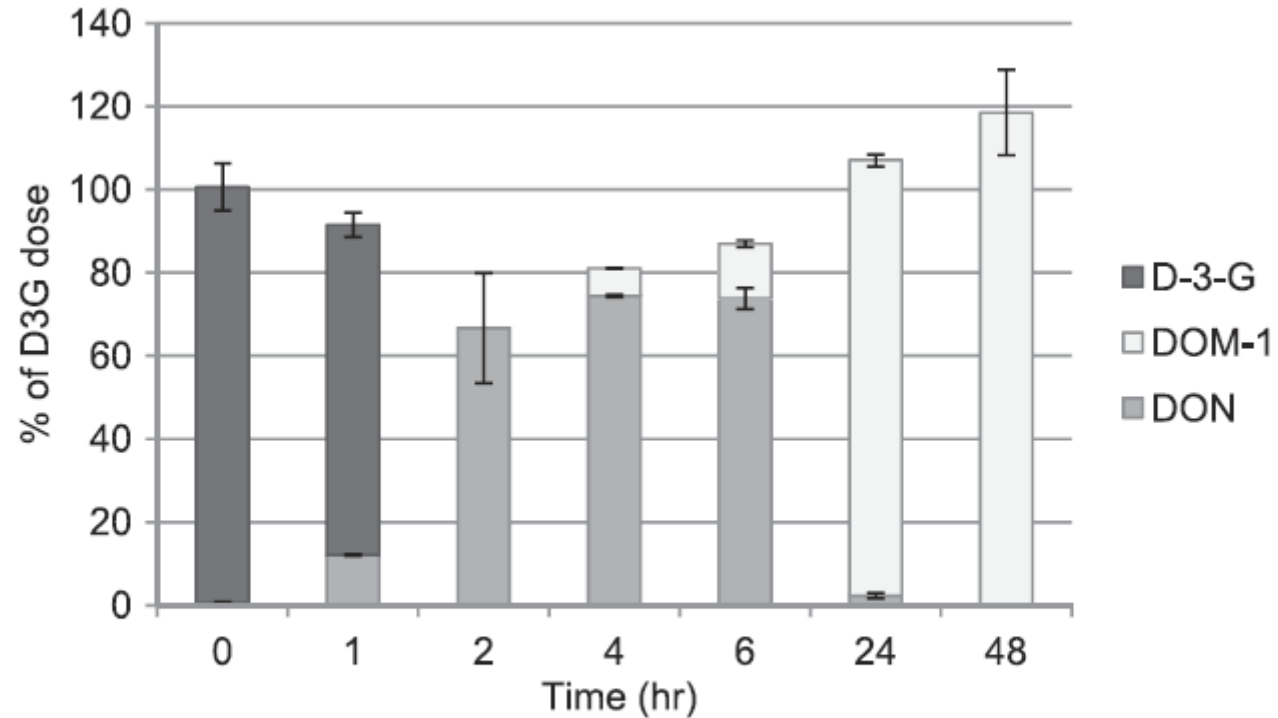
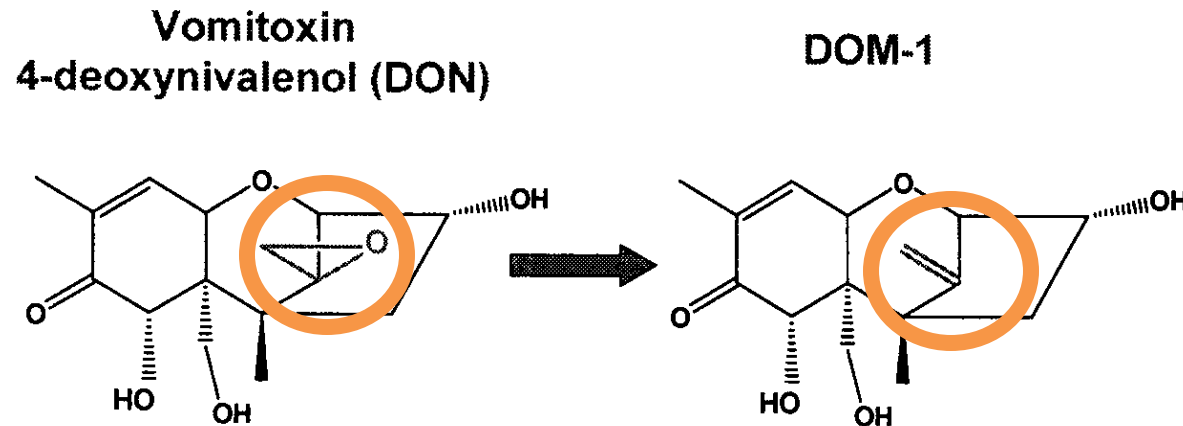


FIG 2 Metabolism of D3G by the human microbiota of volunteer 5. Results are presented as means of duplicate incubations for each time point.

De-epoxy deoxynivalenol (DOM-1)

- Bacterial conversion
- Large intestine
- Epoxide reductase
- Less toxic than DON



Reaction mechanism of the formation of deepoxy-deoxynivalenol from deoxynivalenol.

D3G Metabolism by Human Microbiota

During Incubation

- D3G is degraded
- DON is released
- DON is detoxified to deepoxy-deoxynivalenol (DOM-1)

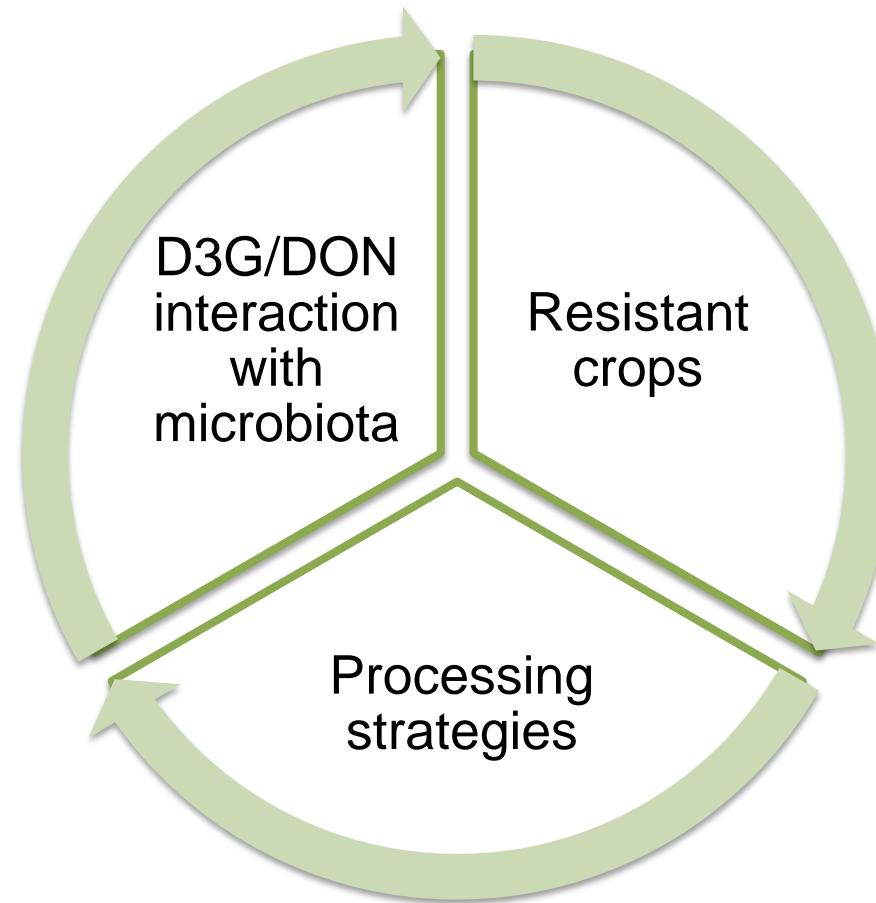
Gratz et al., 2013

D3G and Gut Microbiota

Bacterial hydrolysis of deoxynivalenol-3-glucoside.

Bacterial strain	Hydrolysis after 4 h (%)	Hydrolysis after 8 h (%)
<i>Lactobacillus plantarum</i>	34	62
<i>Enterococcus mundtii</i>	18	38
<i>Enterobacter cloacae</i>	17	25
<i>Bifidobacterium adolescentis</i>	5-17	17-25

Future Research



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- Gardiner, D. M., Osborne, S., Kazan, K., and Manners, J. M. 2009. Low pH regulates the production of deoxynivalenol by *Fusarium graminearum*. Microbiol. 155:3149-3156.
- Government of Saskatchewan. Fusarium head blight. <https://www.saskatchewan.ca/business/agriculture-natural-resources-and-industry/agribusiness-farmers-and-ranchers/crops-and-irrigation/crop-protection/disease/fusarium-head-blight> (accessed 2017 December 17).
- Jin, Z., Zhou, B., Gillespie, J., Gross, T., Barr, J., Simsek, S., Brueggeman, R., and Schwarz, P. 2018. Production of deoxynivalenol (DON) and DON-3-glucoside during the malting of *Fusarium* infected hard red spring wheat. Food Control. 85:6-10.
- Ohio State University Extension. *Fusarium* head blight or head scab of wheat, barley and other small grain crops. <https://ohioline.osu.edu/factsheet/plpath-cer-06>.
- Pestka, J. J., and Smolinski, A. T. 2005. Deoxynivalenol: Toxicology and potential effects on humans. J. Toxicol. Environ. Health, Part B. 8:39-69.
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- U.S. Food & Drug Administration. 2017. Guidance for industry and FDA: Advisory levels for deoxynivalenol (DON) in finished wheat products for human consumption and grains and grain by-products used for animal feed. <https://www.fda.gov/Food/GuidanceRegulation/GuidanceDocumentsRegulatoryInformation/ChemicalContaminantsMetalsNaturalToxinsPesticides/ucm120184.htm> (accessed 2017 December 17).
- Wu, L., Qiu, L., Zhang, H., Sun, J., Hu, X., and Wang, B. 2017. Optimization for the production of deoxynivalenol and zearalenone by *Fusarium graminearum* using response surface methodology. Toxins. 9:57.



muchos gracias!

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